

Restriction Map and Multiple Cloning Site (MCS) of pEYFP-N1. Unique restriction sites are in bold. The Xba I site (*) is methylated in the DNA provided by BD Biosciences Clontech. If you wish to digest this vector with this enzyme, you will need to transform the vector into a dam- host and make fresh DNA.

Description:

591

pEYFP-N1 encodes an enhanced yellow-green variant of the Aequorea victoria green fluorescent protein (GFP). The EYFP gene contains four amino acid substitutions previously published as GFP-10C (1). The fluorescence excitation maximum of EYFP is 513 nm, and the emission spectrum has a peak at 527 nm (in the yellow-green region). When excited at 513 nm, the E_m of EYFP is 36,500 cm⁻¹M⁻¹ and the fluorescence quantum yield is 0.63 (1), resulting in a bright fluorescent signal. The fluorescence level observed from EYFP is roughly equivalent to that from EGFP.

In addition to the chromophore mutations, EYFP contains >190 silent mutations that create an open reading frame comprised almost entirely of preferred human codons (2). Furthermore, upstream sequences flanking EYFP have been converted to a Kozak consensus translation initiation site (3). These changes increase the translational efficiency of the EYFP mRNA and consequently the expression of EYFP in mammalian and plant cells.

The MCS in pEYFP-N1 is between the immediate early promoter of CMV (P_CMVIE) and the EYFP coding sequences. Genes cloned into the MCS will be expressed as fusions to the N-terminus of EYFP if they are in the same reading frame as EYFP and there are no intervening stop codons. The inserted gene should include an initiating ATG codon. EYFP with N-terminal fusion moieties retains the fluorescent properties of the native protein and thus can be used to localize fusion proteins in vivo.

The vector contains an SV40 origin of replication and a neomycin resistance (Neo^r) gene for selection (using G418) in mammalian cells. A bacterial promoter upstream of this cassette (P) expresses kanamycin resistance in E. coli. The vector backbone also provides a pUC19 origin of replication for propagation in E. coli and an f1 origin for single-stranded DNA production.

The recombinant EYFP vector can be transfected into mammalian cells using any standard transfection method. If required, stable transformants can be selected using G418 (4), pEYFP-N1 can also be used simply to express EYFP in a cell line of interest (e.g., as a transfection marker). EGFP, EYFP, and EBFP variants can be used independently or in combination for flow cytometry analysis.

(PR29945; published 03 Octobers 2002)



United States/Canada 800.662.2566 Asia Pacific +1.650.919.7300 Europe +33.(0)1.3904.6880 Japan +81.(0)77.543.6116

Clontech Laboratories, Inc. ATakara Bio Company 1290 Terra Bella Ave. Mountain View, CA 94043 Technical Support (US) E-mail: tech@clontech.com www.clontech.com

Location of features:

- Human cytomegalovirus (CMV) immediate early promoter: 1–589 Enhancer region: 59–465 TATA box: 554–560 Transcription start point: 583 C→G mutation to remove Sac I site: 569
- MCS: 591-671
- Enhanced yellow fluorescent protein (EYFP) gene Kozak consensus translation initiation site: 672–682 Start codon (ATG): 679–681; stop codon: 1396–1398 Insertion of Val at position 2: 682–684 GFP-10C mutations (Ser-65 to Gly: 874–876; Val-68 to Leu: 883–885; Ser-72 to Ala: 895–897; Thr-203 to Tyr: 1288–1290)
- His-231 to Leu mutation (A \rightarrow T): 1373
- SV40 early mRNA polyadenylation signal Polyadenylation signals: 1552–1557 & 1581–1586 mRNA 3' ends: 1590 & 1602
- f1 single-strand DNA origin: 1649–2104 (Packages the noncoding strand of EYFP.)
- Bacterial promoter for expression of Kan^r gene: –35 region: 2166–2171; –10 region: 2189–2194 Transcription start point: 2201
- SV40 origin of replication: 2445–2580
- SV40 early promoter Enhancer (72-bp tandem repeats): 2278–2349 & 2350–2421 21-bp repeats: 2425–2445, 2446–2466, & 2468–2488 Early promoter element: 2501–2507 Major transcription start points: 2497, 2535, 2541 & 2546
- Kanamycin/neomycin resistance gene Neomycin phosphotransferase coding sequences: Start codon (ATG): 2629–2631; stop codon: 3421–3423 G→A mutation to remove *Pst* I site: 2811 C→A (Arg to Ser) mutation to remove *Bss*H II site: 3157
- Herpes simplex virus (HSV) thymidine kinase (TK) polyadenylation signal Polyadenylation signals: 3659–3664 & 3672–3677
- pUC plasmid replication origin: 4008-4651

Primer Locations:

- EGFP-N Sequencing Primer (#6479-1): 745-724
- EGFP-C Sequencing Primer (#6478-1): 1332-1353

Propagation in *E. coli*:

- Suitable host strains: DH5α, HB101, and other general-purpose strains. Single-stranded DNA production requires a host containing an F plasmid such as JM101 or XL1-Blue.
- Selectable marker: plasmid confers resistance to kanamycin (30 µg/ml) in *E. coli* hosts.
- E. coli replication origin: pUC
- Copy number: ~500
- Plasmid incompatibility group: pMB1/CoIE1

References:

- 1. Ormö, M. et al. (1996) Science 273:1392–1395.
- 2. Haas, J., et al. (1996) Curr. Biol. 6:315–324.
- 3. Kozak, M. (1987) Nucleic Acids Res. 15:8125–8148.
- 4. Gorman, C. (1985) In DNA cloning: a practical approach, vol. II. Ed. D. M. Glover. (IRL Press, Oxford, U.K.), pp. 143–190.

Note: The attached sequence file has been compiled from information in the sequence databases, published literature, and other sources, together with partial sequences obtained by BD Biosciences Clontech. This vector has not been completely sequenced.

Notice to Purchaser

Use of BD Biosciences Clontech's Living Colors[™] products containing DNA sequences coding for mutant *Aequorea victoria* green fluorescent protein (GFP) variants or proteins thereof requires a license from Amersham Biosciences under U.S. Patent Nos. 5,625,048; 5,777,079; 6,054,321 and other pending U.S. and foreign patent applications. In addition, certain BD Biosciences Clontech products are made under U.S. Patent No. 5,804,387 licensed from Stanford University.

Not-For-Profit research institutes or entities are granted an automatic license with the purchase of this product for use in non-commercial internal research purposes, the terms of which are disclosed in detail in the license that accompanies the shipment of this product. Such license specifically excludes the right to sell or otherwise transfer this product or its components to third parties.

For-Profit research institutes or entities must obtain a license from Amersham Biosciences. E-mail: gfp@amershambiosciences.com

Please contact BD Biosciences Clontech directly for any other assistance, including purchasing and technical support. All companies and institutions purchasing Living Colors[™] products will be included in a quarterly report to Aurora Biosciences, as required by the BD Biosciences Clontech/Aurora Biosciences license agreement.

This product is intended to be used for research purposes only. It is not to be used for drug or diagnostic purposes nor is it intended for human use. BD Biosciences Clontech products may not be resold, modified for resale, or used to manufacture commercial products without written approval of BD Biosciences Clontech.

© 2002, Becton, Dickinson and Company