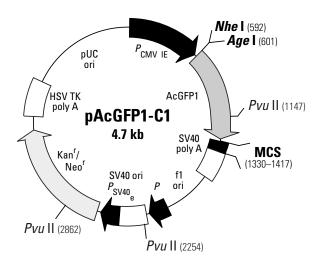
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Restriction Map and Multiple Cloning Site (MCS) of pAcGFP1-C1. Restriction sites shown in bold are unique. The *Xba* I and *BcI* I sites (*) are methylated in the DNA provided by Clontech Laboratories, Inc. . If you wish to digest the vector with these enzymes, you will need to transform the vector into a *dam*⁻ host and make fresh DNA.

Description

pAcGFP1-C1 encodes a green fluorescent protein (GFP) from *Aequorea coerulescens* (excitation maximum = 475 nm; emission maximum = 505 nm). Sequences flanking AcGFP1 have been converted to a Kozak consensus translation initiation site (1) to further increase the translation efficiency in eukaryotic cells. The MCS in pAcGFP1-C1 is downstream of the AcGFP1 coding region, allowing the contruction of a C-terminal fusion protein with the AcGFP1 when genes are cloned in the same reading frame as AcGFP1 and there are no intervening stop codons. SV40 polyadenylation signals downstream of the AcGFP1 gene direct proper processing of the 3' end of the AcGFP1 mRNA. The vector backbone also contains an SV40 origin for replication in mammalian cells expressing the SV40 T-antigen. A neomycin resistance cassette (Neo'), consisting of the SV40 early promoter, the neomycin/kanamycin resistance gene of Tn5, and polyadenylation signals from the Herpes simplex virus thymidine kinase (HSV TK) gene, allows stably transfected eukaryotic cells to be selected using G418. A bacterial promoter upstream of the gene expresses kanamycin resistance in *E. coli*. The pAcGFP1-C1 backbone also provides a pUC origin of replication for propagation in *E. coli* and an f1 origin for single-stranded DNA production.

Use

Fusions to the C terminus of AcGFP1 retain the fluorescent properties of the native protein allowing the localization of the fusion protein *in vivo*. The target gene should be cloned into pAcGFP1-C1 so that it is in frame with the AcGFP1 coding sequences, with no intervening in-frame stop codons. The recombinant AcGFP1 vector can be transfected into mammalian cells using any standard transfection method. If required, stable transformants can be selected using G418 (2). pAcGFP1-C1 can also be used simply to express AcGFP1 in a cell line of interest (e.g., as a transfection marker).



United States/Canada 800.662.2566

Asia Pacific +1.650.919.7300

+1.650.919.7300 **Europe**

+33.(0)1.3904.6880

Japan +81 (0)7

+81.(0)77.543.6116

Clontech Laboratories, Inc. ATakara Bio Company 1290 Terra Bella Ave. Mountain View, CA 94043 Technical Support (US) E-mail: tech@clontech.com www.clontech.com

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pAcGFP-C1 Vector Information

Location of features

• Human cytomegalovirus (CMV) immediate early promoter: 1-589

Enhancer region: 59–465; TATA box: 554–560

Transcription start point: 583

C→G mutation to remove *Sac* I site: 569

 Aequorea coerulescens green fluorescent protein gene Kozak consensus translation initiation site: 606–616

Start codon (ATG): 613-615; Stop codons: 1404-1406, 1408-1410 & 1412-1414

Insertion of Val at position 2: 616-618

Last amino acid: 1327-1329

MCS: 1330–1417

SV40 early mRNA polyadenylation signal

Polyadenylation signals: 1550–1555 & 1579–1584; mRNA 3' ends: 1588 & 1600 f1 single-strand DNA origin: 1647–2102 (Packages the noncoding strand of AcGFP.)

 Bacterial promoter for expression of Kan^r gene –35 region: 2164–2169; –10 region: 2187–2192

Transcription start point: 2199
• SV40 origin of replication: 2443–2578

SV40 early promoter

Enhancer (72-bp tandem repeats): 2276-2347 & 2348-2419

21-bp repeats: 2423-2443, 2444-2464, & 2466-2486

Early promoter element: 2499-2505

Major transcription start points: 2495, 2533, 2539 & 2544

· Kanamycin/neomycin resistance gene

Neomycin phosphotransferase coding sequences:

Start codon (ATG): 2627-2629; stop codon: 3419-3421

G→A mutation to remove Pst I site: 2809

C→A (Arg to Ser) mutation to remove *Bss*H II site: 3155

Herpes simplex virus (HSV) thymidine kinase (TK) polyadenylation signal

Polyadenylation signals: 3657–3662 & 3670–3675

pUC plasmid replication origin: 4006–4649

Propagation in E. coli

• Suitable host strains: DH5α, HB101, and other general purpose strains. Single-stranded DNA production requires a host containing an F plasmid such as JM109 or XL1-Blue.

• Selectable marker: plasmid confers resistance to kanamycin (30 μg/ml) to *E. coli* hosts.

- E. coli replication origin: pUC
- Copy number: ≈500
- · Plasmid incompatibility group: pMB1/ColE1

References

- 1. Kozak, M. (1987) Nucleic Acids Res. 15:8125-8148.
- 2. Gorman, C. (1985) In DNA Cloning: A Practical Approach, Vol. II, Ed. Glover, D. M. (IRL Press, Oxford, UK) pp. 143-190.

Note: The attached sequence file has been compiled from information in the sequence databases, published literature, and other sources, together with partial sequences obtained by Clontech Laboratories, Inc. This vector has not been completely sequenced.

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