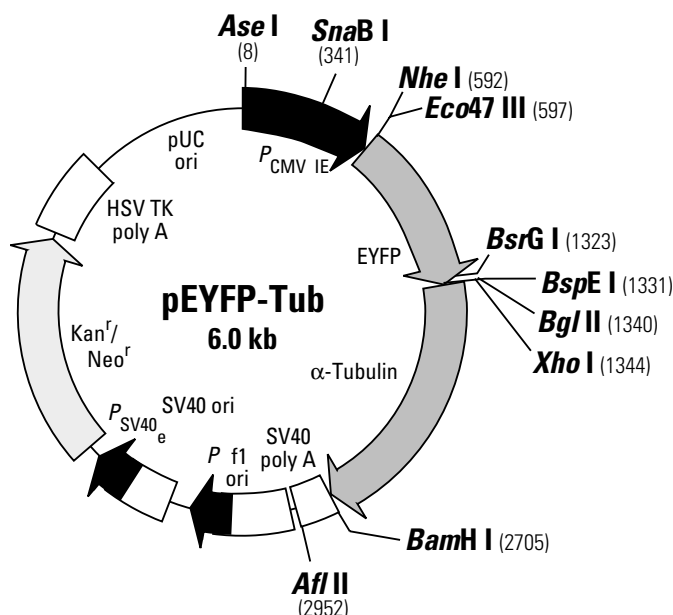




Restriction Map of pEYFP-Tub. All sites shown are unique. The *Xba*I and *Bcl*I sites at positions 2717 & 2727 are methylated in the DNA provided by CLONTECH. If you wish to digest the vector with these enzymes, you will need to transform the vector into a *dam*⁻ host and isolate fresh DNA.

Description

pEYFP-Tub encodes a fusion protein consisting of enhanced yellow fluorescent protein (EYFP) and human α-tubulin (1, 2). EYFP is an enhanced yellow-green variant of the *Aequorea victoria* green fluorescent protein (GFP; 3–5). The EYFP gene contains the four amino acid substitutions previously published as GFP-10C (6): Ser-65 to Gly; Val-68 to Leu; Ser-72 to Ala; and Thr-203 to Tyr. It also contains the Phe-64 to Leu mutation of GFPmut1 (7). The fluorescence excitation maximum of EYFP is 513 nm; the emission spectrum has a peak at 527 nm (in the yellow-green region). When excited at 513 nm, the E_m of EYFP is 36,500 cm⁻¹M⁻¹ and the fluorescence quantum yield is 0.63 (6), resulting in a bright fluorescent signal. The fluorescence observed is roughly equivalent to that from EGFP.

In addition to the chromophore mutations, pEYFP-Tub contains more than 190 silent base changes that correspond to human codon-usage preferences (8). SV40 polyadenylation signals downstream of the EYFP-Tub fusion direct proper processing of the 3' end of the mRNA. The vector backbone also contains an SV40 origin for replication in mammalian cells expressing the SV40 T-antigen. A neomycin resistance cassette (Neo^r) consisting of the SV40 early promoter, the neomycin/kanamycin resistance gene of Tn5, and polyadenylation signals from the herpes simplex virus thymidine kinase (HSV-TK) gene allows stably transfected eukaryotic cells to be selected using G418 (9). A bacterial promoter upstream of this cassette drives expression of the gene encoding kanamycin resistance in *E. coli*. The pEYFP-Tub backbone also provides a pUC origin of replication for propagation in *E. coli* and an f1 origin for single-stranded DNA production.

Use

The pEYFP-Tub Vector is designed for expressing the EYFP-tubulin fusion protein in mammalian cells. pEYFP-Tub can be introduced into mammalian cells using any standard transfection method. If required, stable transformants can be selected using G418 (9). The protein incorporates directly into microtubules and thereby allows the observation of microtubules in living or fixed cells by fluorescence microscopy (10). pEYFP-Tub is not sold as a cloning vector; however, there are unique restriction sites at the 5' end of EYFP, at the EYFP-Tub fusion junction, and at the 3' end of the α-tubulin sequences. Note that the sites at the 3' end of the α-tubulin sequences are downstream of the stop codon.

Location of features

- Human cytomegalovirus (CMV) immediate early promoter: 1–589
Enhancer region: 59–465; TATA box: 554–560
Transcription start point: 583
C→G mutation to remove *Sac* I site: 569
- EYFP-Tubulin fusion: 613–2703
Start codon (ATG): 613–615
Insertion of Val at position 2: 616–618
GFP-10C mutations:
Ser-65 to Gly: 808–810; Val-68 to Leu: 817–819; Ser-72 to Ala: 829–831; Thr-203 to Tyr: 1222–1224
GFPmut1 chromophore mutation (Phe-64 to Leu): 805–808
His-231 to Leu mutation (A→T): 1307
Last amino acid in EYFP: 1327–1329
Human α -tubulin sequence (in frame with EYFP): 1348–2703
Stop codon: 2701–2703
- SV40 early mRNA polyadenylation signal
Polyadenylation signals: 2864–2869 & 2893–2898; mRNA 3' ends: 2902–2914
- f1 single-strand DNA origin: 2961–3416 (packages the noncoding strand of EYFP-Tub)
- Bacterial promoter for expression of Kan^r gene
–35 region: 3478–3483; –10 region: 3501–3506
Transcription start point: 3513
- SV40 origin of replication: 3757–3892
- SV40 early promoter
Enhancer (72-bp tandem repeats): 3590–3661 & 3662–3733
21-bp repeats: 3737–3757, 3758–3778 & 3780–3800
Early promoter element: 3813–3819
Major transcription start points: 3809, 3847, 3853 & 3858
- Kanamycin/neomycin resistance gene
Neomycin phosphotransferase coding sequences:
Start codon (ATG): 3941–3943; stop codon: 4733–4735
G→A mutation to remove *Pst* I site: 4123
C→A (Arg to Ser) mutation to remove *Bss*H II site: 4469
- Herpes simplex virus (HSV) thymidine kinase (TK) polyadenylation signal
Polyadenylation signals: 4971–4976 & 4984–4989
- pUC plasmid replication origin: 5320–5963

Primer Locations

- EGFP-N Sequencing Primer (#6479-1): 679–658; EGFP-C Sequencing Primer (#6478-1): 1266–1287

Propagation in *E. coli*:

- Suitable host strains: DH5 α , HB101, and other general purpose strains. Single-stranded DNA production requires a host containing an F plasmid such as JM109 or XL1-Blue.
- Selectable marker: plasmid confers resistance to kanamycin (30 μ g/ml) to *E. coli* hosts.
- *E. coli* replication origin: pUC; copy number: $\approx$ 500
- Plasmid incompatibility group: pMB1/ColE1

References

1. de Hostos, E., unpublished data.
2. Living Colors Subcellular Localization Vectors (October 1998) *CLONTECHniques* XIII(4):8–9.
3. Prasher, D. C., *et al.* (1992) *Gene* **111**:229–233.
4. Chalfie, M., *et al.* (1994) *Science* **263**:802–805.
5. Inouye, S. & Tsuji, F. I. (1994) *FEBS Letters* **341**:277–280.
6. Ormö, M., *et al.*, (1996) *Science* **273**:1392–1395.
7. Cormack, B., *et al.* (1996) *Gene* **173**:33–38.
8. Haas, J., *et al.* (1996) *Curr. Biol.* **6**:315–324.
9. Gorman, C. (1985) In *DNA Cloning: A Practical Approach*, Vol. II, Ed. Glover, D. M. (IRL Press, Oxford, UK) pp. 143–190.
10. Go to <http://www.stanford.edu/~angelab/> to view time-lapse photos of the microtubule network using EGFP fluorescence.

Notice to Purchaser

Use of CLONTECH's Living Colors® products containing DNA sequences coding for mutant *Aequorea victoria* green fluorescent protein (GFP) variants or proteins thereof requires a license from Aurora Biosciences Corporation under U.S. Patent Nos. 5,625,048; 5,777,079; 6,054,321 and other pending U.S. and foreign patent applications. In addition, certain CLONTECH products are made under U.S. Patent No. 5,804,387 licensed from Stanford University.

Not-For-Profit research institutes or entities are granted an automatic license with the purchase of this product for use in non-commercial internal research purposes, the terms of which are disclosed in detail in the license that accompanies the shipment of this product. Such license specifically excludes the right to sell or otherwise transfer this product or its components to third parties.

For-Profit research institutes or entities that wish to use this product in non-commercial or commercial applications are required to obtain a license from Aurora Biosciences Corporation. For license information contact: Court Turner at 858-404-8416 or Fax 858-404-6743 or www.aurorabio.com. Please contact CLONTECH directly for any other assistance, including purchasing and technical support.

All companies and institutions purchasing Living Colors® products will be included in a quarterly report to Aurora Biosciences Corporation, as required by the CLONTECH/Aurora license agreement.

The attached sequence file has been compiled from information in the sequence databases, published literature, and other sources, together with partial sequences obtained by CLONTECH. This vector has not been completely sequenced.

This product is intended to be used for research purposes only. It is not to be used for drug or diagnostic purposes nor is it intended for human use. CLONTECH products may not be resold, modified for resale, or used to manufacture commercial products without written approval of CLONTECH.

© 2000, CLONTECH Laboratories, Inc.